METABOLISM CLINICAL AND EXPERIMENTAL XX (2015) XXX-XXX



Available online at www.sciencedirect.com

Metabolism

www.metabolismjournal.com



Advanced glycation end products: A link between metabolic and endothelial dysfunction in polycystic ovary syndrome?

Magdalena Pertynska-Marczewska^a, Evanthia Diamanti-Kandarakis^b, John Zhang^c, Zaher Merhi^{d,*}

- ^a Consultant in Obstetrics and Gynecology, 44 Traps Lane, KT3 4SA New Malden, United Kingdom
- ^b Department of Medicine, Endocrine Unit, Medical School University of Athens, Mikras Asias 75, Goudi 115002D27, Athens, Greece
- ^c Reproductive Medicine, New Hope Fertility Center, 4 Columbus Circle, New York, NY, USA
- d Department of Obstetrics and Gynecology, Division of Reproductive Biology, NYU School of Medicine, 180 Varick Street, sixth floor, New York, NY, USA

ARTICLEINFO

Article history: Received 26 April 2015 Accepted 17 August 2015

Keywords:
Polycystic ovary syndrome
Advanced glycation end products
RAGE
Inflammation
Endothelial dysfunction

ABSTRACT

Polycystic ovary syndrome (PCOS), a heterogeneous syndrome of reproductive and metabolic alterations, is associated with increased long-term risk of cardiovascular complications. This phenomenon has been linked to an increase in oxidative stress and inflammatory markers. Advanced glycation end products (AGEs) are pro-inflammatory molecules that trigger a state of intracellular oxidative stress and inflammation after binding to their cell membrane receptors RAGE. The activation of the AGE-RAGE axis has been well known to play a role in atherosclerosis in both men and women. Women with PCOS have systemic chronic inflammatory condition even at the ovarian level as represented by elevated levels of serum/ovarian AGEs and increased expression of the pro-inflammatory RAGE in ovarian tissue. Data also showed the presence of sRAGE in the follicular fluid and its potential protective role against the harmful effect of AGEs on ovarian function. Thus, whether AGE-RAGE axis constitutes a link between metabolic and endothelial dysfunction in women with PCOS is addressed in this review. Additionally, we discuss the role of hormonal changes observed in PCOS and how they are linked with the AGE-RAGE axis in order to better understand the nature of this complex syndrome whose consequences extend well beyond reproduction.

© 2015 Elsevier Inc. All rights reserved.

1. Introduction

Polycystic ovary syndrome (PCOS) is a heterogeneous syndrome that causes reproductive and metabolic alterations

[1,2]. The reported prevalence of PCOS ranges between 2.2% to 26% in various countries, depending on the recruitment process of the study population, and the criteria used for its definition [3–7]. PCOS is associated with hyperandrogenemia

Abbreviations: AGE, advanced glycation end products; RAGE, cellular receptor for AGEs; sRAGE, soluble receptor for AGEs; CVD, cardiovascular disease; PCOS, polycystic ovary syndrome; MG, synthetic methylglyoxal; eNOS, endothelial nitric oxidase synthase NOS; NO, nitric oxidase; ROS, reactive oxygen species; AMH, anti-Mullerian hormone; IVF, in vitro fertilization; TF, tissue factor; TNF- α , tumor necrosis factor alpha; GnRH, gonadotropin-releasing hormone; LH, luteinizing hormone; FSH, follicle-stimulating hormone.

E-mail addresses: m_pertynska@yahoo.com (M. Pertynska-Marczewska), e.diamanti.kandarakis@gmail.com (E. Diamanti-Kandarakis), Johnzhang211@gmail.com (J. Zhang), zom00@hotmail.com (Z. Merhi).

http://dx.doi.org/10.1016/j.metabol.2015.08.010

0026-0495/© 2015 Elsevier Inc. All rights reserved.

^{*} Corresponding author at: NYU School of Medicine, Department of Obstetrics and Gynecology, Division of Reproductive Biology, 180 Varick Street, sixth floor, New York, NY 10014.

[4], insulin resistance (IR) [8,9], impaired glucose tolerance, type 2 diabetes [2], dyslipidemia [10–12], and cardiovascular disease (CVD) [13,14].

Moreover, evidence indicates that low-grade chronic inflammation in PCOS could represent a reason for the long-term metabolic and cardiovascular complications [15]. CVD is one of the leading causes of female morbidity and mortality in the United States [16]. Given its high prevalence, PCOS may potentially account for a significant proportion of atherosclerotic heart disease observed in women [17], independent of obesity or age [18–20]. These findings were reflected by increased coronary calcium, increased carotid intima- media thickness, and endothelial dysfunction [21,22].

One of the emerging pro-inflammatory molecules that are elevated in PCOS is advanced glycation end-products (AGEs). Women with PCOS have elevated serum AGEs and an upregulation of the membranous pro-inflammatory receptor RAGE (receptor for AGEs) was demonstrated in their ovaries [23]. With the increasing evidence for AGEs' role in PCOS-associated inflammation and their role in CVD, this review aims to present an update on the mechanisms by which AGEs act and how the AGE-RAGE axis could potentially play a pivotal role in the pathophysiology of CVD observed in women with PCOS.

2. How do AGEs Form?

AGEs can be formed either endogenously or exogenously [24–26]. Endogenously, advanced glycation takes place in all cell types and refers to a reaction, known as the Maillard reaction, between reducing sugars and amino residues present in proteins, lipids, and DNA. This is followed by rearrangements and final cross-linking to generate AGEs leading to loss of protein structure and function, followed by some instances with cellular apoptosis. Examples of AGEs include N°-(carboxymethyl) lysine (CML), pyrraline and pentosidine [26]. AGEs accumulate in the serum and tissues with aging and their formation is accelerated by hyperglycemia, IR, obesity, metabolic syndrome, hypoxia and oxidative stress [26].

Exogenous AGEs could be introduced into the circulation together with nutrients processed by common methods such as dry heat or other food processing methods for example ionization [27]. Human and animal studies demonstrated that about 10% of AGEs contained in a meal could be absorbed into the circulation, of which two-thirds remain in the body for 72 h [28,29]; a period of time long enough to promote oxidative stress and cause tissue injury [28]. Diamanti-Kandarakis et al. demonstrated that in female rats, a high-AGE diet for 6 months increased serum AGEs' levels and caused higher deposition of their pro-inflammatory RAGE in the ovarian tissue and caused an increase in ovarian weight (usually observed in women with PCOS) compared to animals fed a low-AGE diet [30]. These data provide evidence for the impact of dietary AGEs on reproductive dysfunction, causing a pattern observed in PCOS.

Another exogenous source of AGEs is smoking [31]. Both the aqueous extracts of tobacco and cigarette smoke contain glycotoxins—highly reactive glycation products that can rapidly induce AGE formation on proteins. Smoking has been shown to worsen the already elevated risk for metabolic syndrome in women with PCOS [32]. Whether this phenomenon is partly induced by AGEs remains to be determined [31].

3. Different Types of AGEs' Receptors

AGEs could cause tissue injury via cross-linking with extracellular matrix of tissues throughout the body or by binding to RAGE on the surface of various cells such as endothelial cells and mononuclear phagocytes [33]. RAGE, a member of the immunoglobulin superfamily, has transmembrane, cytosolic and extracellular domains. RAGE is expressed in many tissues including the vascular system [34]. The binding of AGEs to RAGE causes intracellular signaling that leads to reactive oxygen species (ROS) production mainly via activation of the proinflammatory transcription factor (NF-kB) and nicotinamide adenine dinucleotide phosphate (NADPH) oxidase (Fig. 1). Subsequently, there is an increase in the production of proinflammatory cytokines (such as IL-1, IL-6 and IL-8), chemokines, apoptosis regulators such as bcl-2, Fas, adhesion molecules (such as VCAM-1 and ICAM-1), as well as macrophage and platelet activation [35,36]. Interestingly, ROS production caused by RAGE activation causes a positive feedback loop therefore up-regulating RAGE expression [37,38], thus leading to exacerbation of the inflammatory processes. Induction of RAGE has been documented in inflammatory processes, atherosclerosis and recently PCOS [34,39].

The soluble form of RAGE (sRAGE) is a product of both splicing of RAGE gene and/or cleavage of membrane-bound RAGE by proteases belonging to the zinc-dependent metzincin family of metalloproteases [40]. Following its formation, sRAGE receptors circulate and act as decoy for AGEs by binding them and thus competitively inhibiting AGE–RAGE interaction and its downstream pro-inflammatory signaling [41]. In many instances, sRAGE is often considered an anti-inflammatory receptor.

4. The Role of Hormonal Change Observed in PCOS

The metabolic-hormonal complexity observed in PCOS represents a biological model illustrating the relationship between hormonal pattern and cardiovascular risk profile [42,43]. The key endocrine abnormalities of the reproductive axis include accelerated GnRH pulsatile activity, increased secretion of pituitary LH, theca-stromal cell hyperactivity and hypofunction of the FSH-granulosa cell axis [44,45]. Increased LH pulse frequency and amplitude leading to persistently increased LH levels may directly enhance theca androgen synthesis. On the other hand, it has been suggested that elevated LH levels result from an impaired negative feedback on LH secretion, due to excessive androgen action on the hypothalamic-pituitary axis [46]. Insulin may play a part in the development of the typical increased amplitude and frequency of GnRH and LH pulse secretion seen in PCOS since, elevation of LH and GnRH secretion in response to insulin infusion have been observed in vitro, both in dosedependent and in time-dependent fashions [45,47]. The compensatory hyperinsulinemia is considered to be a promoter of the hyperandrogenism and chronic oligo- or

AITHOLL IN THESS

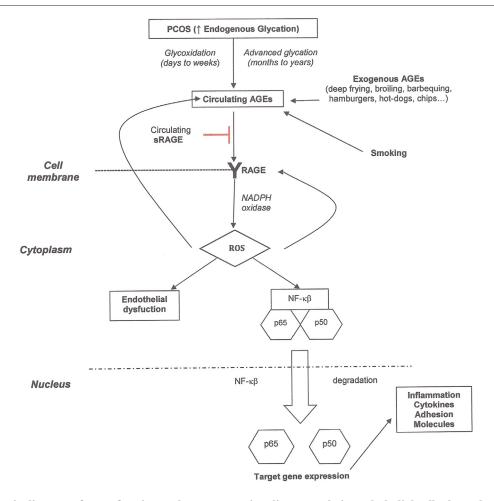


Fig. 1 – A schematic diagram of AGEs forming and AGE-RAGE signaling cascade in endothelial cells through translocation of NFκB into the nucleus.AGEs are formed either *endogenously* or *exogenously*. Intracellular effects are mediated through binding of AGEs to specific cellular receptors RAGE, leading to ROS production via activation of NADPH oxidase with subsequent nuclear translocation of NF-κB, which targets several genes involved in inflammation, immune response, and apoptosis. Additionally, increased superoxide production causes an increased formation of AGEs and over-expression of RAGE, acting as positive-feedback loop and thus worsening the inflammatory response. sRAGE acts as a decoy for ligands, thus competitively inhibiting AGE-RAGE interaction and its downstream pro-inflammatory signaling. AGEs (advanced glycation end products), cellular receptor for AGEs (RAGE), soluble receptor for AGEs (sRAGE), NF- B (nuclear factor kappa-light-chain-enhancer of activated B cells), ROS (reactive oxygen species).

anovulation, particularly when there is concomitant obesity [48–51].

Insulin can also act indirectly to raise the serum concentration of free testosterone, the level of which does not seem to be tightly regulated in women, by inhibiting the hepatic production of sex-hormone–binding globulin (SHBG) [52]. Therefore, an improvement in insulin sensitivity in women with PCOS leads to a decrease in ovarian androgen biosynthesis, an increase in the concentration of SHBG, and a resultant decrease in free testosterone concentration [53]. Interestingly, serum levels of AGEs correlate with the hormonal abnormalities observed in women with PCOS [23,54]; for instance, we have shown a correlation between serum AGEs and serum testosterone [54].

Additionally, it has been reported that changes in dietary AGEs parallel changes in insulin sensitivity, oxidative stress

and hormonal status in women with PCOS and therefore lowering the concentration of AGEs in food may improve these variables [55]. Serum levels of AGEs, testosterone, oxidative stress, insulin and HOMA-IR index were significantly increased on the isocaloric diet with high AGEs compared to the hypocaloric diet with ad-libitum AGEs content diet and subsequently decreased on the isocaloric diet with low AGEs diet (compared to high AGEs) (p < 0.05 for all parameters). BMI remained unaltered throughout the high AGEs and low AGEs periods compared to the hypocaloric diet period. Serum AGEs and dietary AGEs were strongly correlated with insulin during low AGEs period. Such findings may have important clinical implications for women with PCOS, as dietary habits seem to play a role in the pathophysiogy of the syndrome [56]. Additionally, dietary AGEs, regardless of BMI, may modify both metabolic and hormonal parameters of women with PCOS [55]. Moreover, the diet-induced elevation in AGEs' levels and the increased oxidative stress may directly induce insulin-signaling defects [57].

5. What Happens to Circulating sRAGE and Ovarian AGEs and RAGE in PCOS?

Serum AGEs are distinctly elevated in women with PCOS compared to women who only have the isolated characteristics of PCOS syndrome [23]. Diamanti-Kandarakis et al. demonstrated that women with PCOS exhibited statistically higher AGE levels (7.96 \pm 1.87 U/ml, p < 0.001) compared to those with isolated hyperandrogenaemia (5.61 ± 0.61 U/ml), anovulation (5.53 ± 1.06 U/ml), women with PCO morphology only (rather than PCOS) on ultrasound (5.26 \pm 0.25 U/ml) and controls $(5.86 \pm 0.89 \text{ U/ml})$ [23]. Moreover, compared to women without PCOS, insulin resistant women with PCOS (without overt hyperglycemia) have higher levels of serum AGEs (9.81 \pm 0.16 vs. 5.11 \pm 0.16, p < 0.0001), and up-regulation of RAGE expression in their monocytes (30.91 ± 10.11 vs. 7.97 ± 2.61 , p < 0.02) [54]. Additionally, IR indices such as Quantitative Insulin Sensitivity Check Index (QUICKI) and Homeostasis Model Assessment (HOMA) showed a positive correlation with serum AGE levels [54].

Interestingly, phosphatidyl inositol-3 kinase (PI-3 K) mediates insulin signaling at the post-receptor level and it also mediates the clearance of AGEs via the macrophage scavenger receptor (MSR) pathway [58,59]. Therefore, the presence of IR in normoglycemic women with PCOS could alter the clearance of AGEs via decreased activity of PI-3 K, which has been demonstrated previously in women with PCOS [60,61]. Hence, one could conclude that PCOS is associated with elevated levels of AGEs that, in turn, can exacerbate manifestations of PCOS such as IR.

As mentioned earlier, data to date demonstrated that women with PCOS have ovarian chronic inflammatory condition represented by elevated levels of serum/ovarian AGEs and up-regulation of ovarian RAGE [23,33]. Diamanti-Kandarakis et al. demonstrated that in PCOS ovaries, granulosa cells displayed stronger RAGE expression then theca interna cells when compared to controls [33]. The authors also showed that NF-κB (p50/p65) was expressed in the cytoplasm of theca interna and granulosa cells of both normal and PCOS ovaries; whereas the NF-κB p65 subunit was only observed in granulosa cells nuclei of PCOS tissue [33]. Interestingly, it was suggested that accumulation of AGEs' products at the level of the ovarian follicle might trigger early ovarian aging [62] or could be responsible for reduced glucose uptake by granulosa cells, potentially altering follicular growth [26]. Additionally, very recently Diamanti-Kandarakis et al. demonstrated that AGEs interfere with insulin signaling in granulosa cells and prevent glucose transporter (Glut-4) membrane translocation. These data suggest that intra-ovarian AGEs accumulation may contribute to the pathophysiology of conditions associated with anovulation and insulin resistance such as PCOS [63]. Presently, there are no data comparing levels of sRAGE between women with or without PCOS. Anti-Mullerian hormone (AMH) is considered one of the best markers of ovarian reserve. Interestingly, Merhi et al. showed a positive correlation

between follicular fluid sRAGE and follicular fluid AMH protein levels [64]. Moreover, it was demonstrated that follicular fluid sRAGE protein concentration, similar to AMH, could predict the number of oocytes retrieved for in vitro fertilization. The authors concluded that sRAGE may relate to the reproductive environment and therefore to reproductive potential. Specifically this may indicate a functionality role for sRAGE in the follicular fluid and the activity of the AGE–RAGE system in ovarian follicles [64] implying a protective role for sRAGE from the harmful effect of AGEs on ovarian folliculogenesis [65].

6. Do AGEs Exacerbate Insulin Resistance in PCOS?

It is well established that almost two thirds of women with PCOS are predisposed to developing IR [66] and ultimately diabetes [67–69], both of which are typically exacerbated by obesity [70,71]. AGEs have been implicated in the development of IR in PCOS (Fig. 2) [72]. This aspect has been discussed in details in our previous paper [65]. Additionally, restriction of AGEs, without altering caloric or nutrient intake, has been shown to ameliorate IR and extend mice life span [73]. Moreover, in patients with type 2 diabetes, AGE-restricted diet improved IR [74]. A reduced demand on beta cells for insulin release appears to be directly related to AGE restriction [74]. A study in overweight women demonstrated that consumption of a low-AGE diet over 4 weeks improved IR [75].

An interesting observation was made in that by the fifth generation, methylglyoxal-fed F5-mice developed insulin resistance followed by diabetes mellitus at 16-18 months of age, whereas AGE-restricted F5-mice did not develop these changes until the age of 36 months or more [76]. The changes represented the effect of a known AGE (methylglyoxal) added to food that otherwise had a low AGE content [76]. Metformin therapy is known to improve the metabolic and hormonal profiles in PCOS [77,78]. In one study [77], women with PCOS received a dose of 1700 mg of metformin daily for a 6-month period. Serum levels of AGEs were reduced after metformin administration in 22 women with PCOS (9.98 \pm 0.13 [before metformin] vs. 9.86 \pm 0.11 [after metformin], p < 0.05). In a subgroup analysis of 16 women with PCOS and normal glucose tolerance, the drop in serum AGEs was potentiated (9.98 \pm 0.19 [before] vs. 9.81 \pm 0.15 [after], p < 0.02). Body mass index remained unchanged after metformin therapy but there was a drop in testosterone levels (p = 0.01) and free androgen index (p = 0.009) [77]. That study indicated that metformin therapy reduces the levels of the atherogenic AGE molecules in the serum of women with PCOS. Besides metformin, a 6-month treatment with a combination of Nacetylcysteine (1200 mg) plus L-arginine (ARG) (1600 mg) showed a potential to restore gonadal function and to improve insulin sensitivity in women PCOS [79].

7. Are AGEs Associated With Adiposity in PCOS?

Approximately 30% to 75% of women with PCOS are obese [80]. Jia et al. demonstrated a direct link between AGEs and adiposity where MG directly stimulated adipogenesis in a cell line model derived from adipose tissue [81]. We have previously

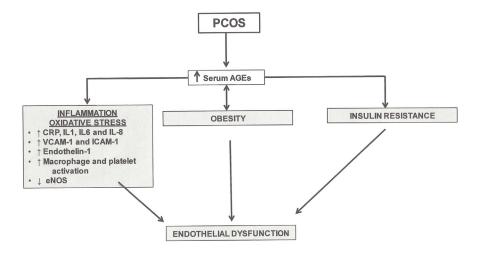


Fig. 2 – Increased serum levels of AGEs could represent a trigger for endothelial cell dysfunction in women with PCOS. Serum levels of AGEs are elevated in PCOS. Elevation in serum AGEs is associated with obesity, insulin resistance, and inflammation; all of which are triggers for endothelial cell dysfunction. IL (interleukin), ICAM-1 (intercellular adhesion molecule 1), VCAM-1 (vascular cell adhesion molecule 1), eNOS (endothelial nitric oxide synthases).

discussed that aspect, as well as sRAGE correlation with adiponectin, in details in a previous paper [65]. Very recently, more data demonstrated that high AGEs may represent a risk factor for metabolic syndrome, type 2 diabetes, and cardiovascular disease [82]. In a study that aimed to assess the association of endogenous and exogenous AGEs with metabolic syndrome criteria, AGEs (*N-carboxymethyllysine, methylglyoxal) were markedly elevated in obese persons with more than one other metabolic syndrome criteria but not in obese without metabolic syndrome criteria. Therefore, high dietary AGE consumption and serum AGEs' levels may link healthy obesity to at-risk obesity [82].

Dietary AGEs may modify both metabolic and hormonal parameters of affected women, without changes in BMI [55]. Moreover, diet-induced elevation of AGEs' levels and the increased oxidative stress may directly induce insulin-signaling defects, as suggested by previous studies [55,57]. Short- and long-term food restriction has been shown to reduce the accumulation of AGEs [83,84]. Additionally, Gaen et al. showed that RAGE-mediated CML (one of the AGEs) accumulation in human adipose tissue and the activation of the CML-RAGE axis caused dysregulation of adipokines in obesity [85]. Further, RAGE deficiency was associated with decreased fat mass and smaller adipocyte size, less body weight, and less epidermal fat weight [86,87]. Several studies demonstrated that plasma sRAGE level is inversely correlated with obesity [88,89]. Interestingly, sRAGE levels rise significantly after surgical weight loss [90], further suggesting an association between AGEs and adiposity (Fig. 2). Diet modification has been shown to alter PCOS phenotypes; for instance, polyunsaturated fatty acids (PUFAs) modulated hormonal and lipid profiles and supplementation with long-chain (LC) n-3 (omega-3) PUFAs improved androgenic profiles in women with PCOS where LC n-3 PUFA supplementation reduced plasma bioavailable testosterone concentrations [91]. Additionally, Omega-3 fatty acids have been shown to improve insulin sensitivity in PCOS patients [92].

8. Do AGEs Cause Endothelial and Vascular Dysfunction in Women With PCOS?

Women with PCOS have a higher prevalence of subclinical atherosclerosis, as reflected in dysregulation of endothelial function assessed by decreased flow mediated dilation, increased carotid intimal-medial thickness (CIMT), and presence of coronary artery calcification [93–96]. However, in the study population that consisted of 85 Taiwanese–Chinese women (aged 17–35 years), there were no significant differences in CIMT between the PCOS and the controls groups [97]. Whether the disagreement between these studies was due to ethnic difference in the population studied remains to be determined.

Moreover, it has been shown that endothelial function is compromised even in young and nonobese women with PCOS [98,99]. Interestingly, a recent study demonstrated that women with PCOS have increased sympathetic drive and impaired endothelial function independent of obesity and metabolic disturbances [100]. That was demonstrated by an elevated multiunit muscle sympathetic nervous system activity and elevated firing rate of the vasoconstrictor neurons in women with PCOS [100].

Endothelial dysfunction is promoted by high levels of IL-6 and TNF- α and low expression/activity of endothelial nitric oxide synthase (eNOS), which lead to decreased nitric oxide (NO) synthesis and reduced vasodilation [101]. An attenuation of NO-based signaling in platelets and blood vessels has been found in women with PCOS and these changes were present from early adult life, contributing to premature atherogenesis [102]. Additionally, several studies have shown that PCOS is associated with high levels of adhesion molecules, such as soluble intercellular adhesion molecule 1, soluble vascular adhesion molecule 1; endothelin-1, and sE-selectin—all of which are known markers of endothelial dysfunction [103,104]. Endothelin-1 (ET-1) (a peptide which causes endothelial dysfunction) levels were found to be

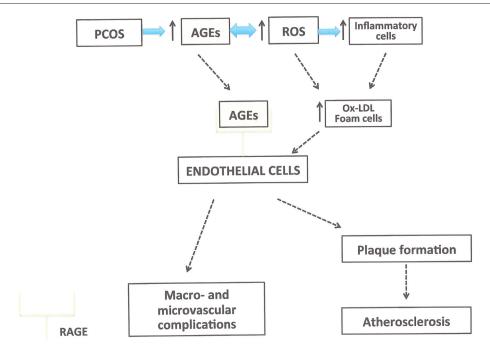


Fig. 3 – AGEs and RAGE play a central role in the development of macro- and microvascular complications in women with PCOS.AGE-RAGE binding elicits reactive oxygen species (ROS) generation that leads to the formation of foam cells causing atherosclerosis, macro- and microvascular complications.Ox-LDL (oxidized low-density lipoprotein).

positively and strongly associated with AGEs in both women with PCOS and controls, suggesting that the detrimental effect of AGEs on endothelial cells may involve increased ET-1 production. Since ET-1 has been shown to enhance oxidative stress, it could possibly be creating a vicious cycle between ET-1 and AGEs in PCOS (Fig. 2) [105]. Interestingly, in a study where women with PCOS had elevated levels of ET-1 and lower levels of NO compared to controls of similar age and BMI, metformin administration reduced ET-1 and increased NO levels in PCOS [106].

Several authors have demonstrated that sRAGE levels were lower in subjects with atheromatous plaques than those without a plaque, whereas serum AGEs levels were significantly higher [107–109]. Interestingly, AGEs, TNF- α , and estradiol (E2) all upregulate RAGE mRNA and protein levels in human vascular endothelial cells, and this process was mediated by two distinct nuclear complexes: p65/p50 NF- κ B and Sp-1/ER α [110]. Therefore, continuous exposure to AGE, TNF- α , and/or E2 and sustained enhancement of RAGE expression may cause a further accumulation of AGE in the vasculature, resulting in an exacerbation of AGE–RAGE mediated vascular dysfunction (Fig. 3).

Interestingly, data reported an association between polymorphism in the TNF- α gene promoter and PCOS [111]. Analysis demonstrated that the –308A variant in the promoter of the TNF α gene clearly influenced the phenotype, resulting in increased basal and leuprolide-stimulated androgen concentrations in the group of carriers of this variant [111]. Additionally, data showed that in a Korean population, the –1031(T/C) polymorphism in the promoter region of TNF- α gene was associated with PCOS [112].

Osteoprotegerin is a cytokine receptor and a member of the TNF receptor superfamily, with anti-inflammatory and antiapoptotic effects [113]. In PCOS, osteoprotegerin has been negatively related with the amount of circulating androgens, while it is simultaneously associated with endothelial dysfunction and IR, independent of adiposity and androgen excess [113,114].

Asymmetrical dimethylarginine (ADMA), a guanidinosubstituted analog of L-arginine, is a potent endogenous competitive inhibitor of the endothelial NO synthase [115]. Serum levels of ADMA have been shown to be elevated in different pathological conditions such as hypertension, diabetes, renal disease, and dyslipidemia [116-119]. Increased levels of ADMA reduce NO formation and are associated with endothelial dysfunction, thus ADMA has been regarded as a novel cardiovascular risk factor [115,120,121]. Insulin resistance may be the underlying mechanism of endothelial dysfunction through NO pathway in PCOS [122]. One study demonstrated that women with PCOS have elevated ADMA levels suggesting an increased risk for an early onset CVD [123]. In that study, women with PCOS (n = 83) were compared with a control group of healthy women (n = 39). The results showed that ADMA levels were significantly higher (0.56 ± 0.15 vs. 0.50 \pm 0.10; p = 0.024; remained statistically significant after adjustment for BMI) and the arginine to ADMA ratio was lower (159.9 \pm 43.6 vs. 195.1 \pm 74.7; p = 0.028) in women with PCOS as compared to the control group [123]. In women with PCOS, ADMA plasma concentrations were positively correlated with BMI (r = 0.353; p = 0.001), fasting insulin (r = 0.405; p < 0.001), and fasting C peptide (r = 0.348; p = 0.001) [123]. Interestingly, ADMA levels decreased significantly after 6 months treatment with metformin (0.53 \pm 0.06 vs. 0.46 ± 0.09 ; p = 0.013; n = 21) [123]. This well-designed study supports the hypothesis that ADMA could represent an important link between impaired insulin action and endothelial dysfunction in women with PCOS.

9. Conclusion

PCOS, one of the most common endocrine disorders in women, increases the risk of CVD. Interestingly, the proatherogenic inflammatory molecules AGEs are elevated in the serum of women with PCOS. AGEs have been linked to the metabolic and reproductive disturbances observed in PCOS and there are data suggesting that blockade of the pro-inflammatory cellular receptors RAGE might be beneficial in IR states and oxidative stress conditions.

Recognizing the impact of AGEs on atherosclerosis, pharmacological strategies targeting reduction in the pathophysiological mechanisms caused by AGEs are being developed: 1) the blockade of AGEs' formation, 2) the prevention of AGE-RAGE interaction, and 3) the suppression of the RAGE downstream pathway, represent promising therapeutic strategies for CVD. The application of these promising therapies in women with PCOS has the potential to alleviate the long-term cardiovascular complications observed in this patient population. This review reinforces the need for more research pertaining to slowing down the AGE-RAGE axis in women with PCOS in order to alleviate the metabolic and endothelial dysfunctions that cause serious health consequences in this patient population.

Authors' Contribution

M.P-M proposed the idea and wrote the first draft of the manuscript.

M.P-M and Z.M. did literature review and wrote the outline and the first draft of the manuscript.

J.Z. drafted Figs. 2 and 3 along with their legends, and edited sections of the manuscript.

E.D-K edited the manuscript and provided critical revisions to the manuscript.

We do not have any potential conflict of interest to disclose.

Acknowledgments

Grant from "Ferring Pharmaceuticals Inc" to Z.M.

REFERENCES

- [1] Repaci A, Gambineri A, Pasquali R. The role of low-grade inflammation in the polycystic ovary syndrome. Mol Cell Endocrinol 2011;335:30–41.
- [2] Moran LJ, Strauss BJ, Teede HJ. Diabetes risk score in the diagnostic categories of polycystic ovary syndrome. Fertil Steril 2011;95:1742–8.
- [3] Tehrani FR, Simbar M, Tohidi M, Hosseinpanah F, Azizi F. The prevalence of polycystic ovary syndrome in a community sample of Iranian population: Iranian PCOS prevalence study. Reprod Biol Endocrinol 2011;9:39–45.
- [4] Azziz R, Woods KS, Reyna R, Key TJ, Knochenhauer ES, Yildiz BO. The prevalence and features of the polycystic ovary syndrome in an unselected population. J Clin Endocrinol Metab 2004;89: 2745–9.

- [5] March WA, Moore VM, Willson KJ, Phillips DI, Norman RJ, Davies MJ. The prevalence of polycystic ovary syndrome in a community sample assessed under contrasting diagnostic criteria. Hum Reprod 2010;25:544–51.
- [6] Azziz R. PCOS: a diagnostic challenge. Reprod Biomed Online 2004;8:644–8.
- [7] Diamanti-Kandarakis E, Kouli CR, Bergiele AT, Filandra FA, Tsianateli TC, Spina GG, et al. A survey of the polycystic ovary syndrome in the Greek island of Lesbos: hormonal and metabolic profile. J Clin Endocrinol Metab 1999;84:4006–11.
- [8] Carmina E, Lobo RA. Use of fasting blood to assess the prevalence of insulin resistance in women with polycystic ovary syndrome. Fertil Steril 2004;82:661–5.
- [9] Cussons AJ, Stuckey BG, Watts GF. Cardiovascular disease in the polycystic ovary syndrome: new insights and perspectives. Atherosclerosis 2006;185:227–39.
- [10] Berneis K, Rizzo M, Hersberger M, Rini GB, Di Fede G, Pepe I, et al. Atherogenic forms of dyslipidaemia in women with polycystic ovary syndrome. Int J Clin Pract 2009;63:56–62.
- [11] Rizzo M, Berneis K. Low-density lipoprotein size and cardiovascular risk assessment. QJM 2006;99:1–14.
- [12] Valkenburg O, Steegers-Theunissen RP, Smedts HP, Dallinga-Thie GM, Fauser BC, Westerveld EH, et al. A more atherogenic serum lipoprotein profile is present in women with polycystic ovary syndrome: a case-control study. J Clin Endocrinol Metab 2008;93:470–6.
- [13] Guzick DS. Cardiovascular risk in women with polycystic ovarian syndrome. Semin Reprod Endocrinol 1996;14:45–9.
- [14] Wild RA, Carmina E, Diamanti-Kandarakis E, Dokras A, Escobar-Morreale HF, Futterweit W, et al. Assessment of cardiovascular risk and prevention of cardiovascular disease in women with the polycystic ovary syndrome: a consensus statement by the Androgen Excess and Polycystic Ovary Syndrome (AE-PCOS) Society. J Clin Endocrinol Metab 2010;95:2038–49.
- [15] Benson S, Janssen OE, Hahn S, Tan S, Dietz T, Mann K, et al. Obesity, depression, and chronic low-grade inflammation in women with polycystic ovary syndrome. Brain Behav Immun 2008;22:177–84.
- [16] Heron MP, Smith BL. Deaths: leading causes for 2003. Natl Vital Stat Rep 2007;55:1–92.
- [17] Dokras A. Cardiovascular disease risk factors in polycystic ovary syndrome. Semin Reprod Med 2008;26:39–44.
- [18] Shroff R, Kerchner A, Maifeld M, Van Beek EJ, Jagasia D, Dokras A. Young obese women with polycystic ovary syndrome have evidence of early coronary atherosclerosis. J Clin Endocrinol Metab 2007;92:4609–14.
- [19] Birdsall MA, Farquhar CM, White HD. Association between polycystic ovaries and extent of coronary artery disease in women having cardiac catheterization. Ann Intern Med 1997;126:32–5.
- [20] Wild RA, Grubb B, Hartz A, Van Nort JJ, Bachman W, Bartholomew M. Clinical signs of androgen excess as risk factors for coronary artery disease. Fertil Steril 1990;54:255–9.
- [21] Guzick DS, Talbott EO, Sutton-Tyrrell K, Herzog HC, Kuller LH, Wolfson Jr SK. Carotid atherosclerosis in women with polycystic ovary syndrome: initial results from a case-control study. Am J Obstet Gynecol 1996;174:1224–9 [discussion 9–32].
- [22] Talbott EO, Guzick DS, Sutton-Tyrrell K, McHugh-Pemu KP, Zborowski JV, Remsberg KE, et al. Evidence for association between polycystic ovary syndrome and premature carotid atherosclerosis in middle-aged women. Arterioscler Thromb Vasc Biol 2000;20:2414–21.
- [23] Diamanti-Kandarakis E, Katsikis I, Piperi C, Kandaraki E, Piouka A, Papavassiliou AG, et al. Increased serum advanced glycation endproducts is a distinct finding in lean women with polycystic ovary syndrome (PCOS). Clin Endocrinol (Oxf) 2008;69:634–41.
- [24] Uribarri J, Cai W, Peppa M, Goodman S, Ferrucci L, Striker G, et al. Circulating glycotoxins and dietary advanced glycation

- endproducts: two links to inflammatory response, oxidative stress, and aging. J Gerontol A: Biol Med Sci 2007;62:427–33.
- [25] Zhang L, Bukulin M, Kojro E, Roth A, Metz VV, Fahrenholz F, et al. Receptor for advanced glycation end products is subjected to protein ectodomain shedding by metalloproteinases. J Biol Chem 2008;283:35507–16.
- [26] Piperi C, Adamopoulos C, Dalagiorgou G, Diamanti-Kandarakis E, Papavassiliou AG. Crosstalk between advanced glycation and endoplasmic reticulum stress: emerging therapeutic targeting for metabolic diseases. J Clin Endocrinol Metab 2012;97:2231–42.
- [27] Koschinsky T, He CJ, Mitsuhashi T, Bucala R, Liu C, Buenting C, et al. Orally absorbed reactive glycation products (glycotoxins): an environmental risk factor in diabetic nephropathy. Proc Natl Acad Sci U S A 1997;94:6474–9.
- [28] Vlassara H, Uribarri J. Advanced glycation end products (AGE) and diabetes: cause, effect, or both? Curr Diab Rep 2014:14:453–68.
- [29] Vlassara H, Palace MR. Diabetes and advanced glycation endproducts. J Intern Med 2002;251:87–101.
- [30] Diamanti-Kandarakis E, Piperi C, Korkolopoulou P, Kandaraki E, Levidou G, Papalois A, et al. Accumulation of dietary glycotoxins in the reproductive system of normal female rats. J Mol Med 2007;85:1413–20.
- [31] Cerami C, Founds H, Nicholl I, Mitsuhashi T, Giordano D, Vanpatten S, et al. Tobacco smoke is a source of toxic reactive glycation products. Proc Natl Acad Sci U S A 1997;94:13915–20.
- [32] Pau CT, Keefe CC, Welt CK. Cigarette smoking, nicotine levels and increased risk for metabolic syndrome in women with polycystic ovary syndrome. Gynecol Endocrinol 2013;29:551–5.
- [33] Diamanti-Kandarakis E, Piperi C, Patsouris E, Korkolopoulou P, Panidis D, Pawelczyk L, et al. Immunohistochemical localization of advanced glycation end-products (AGEs) and their receptor (RAGE) in polycystic and normal ovaries. Histochem Cell Biol 2007;127:581–9.
- [34] Kalea AZ, Schmidt AM, Hudson BI. RAGE: a novel biological and genetic marker for vascular disease. Clin Sci (Lond) 2009;116:621–37.
- [35] Yan SF, Ramasamy R, Schmidt AM. Receptor for AGE (RAGE) and its ligands-cast into leading roles in diabetes and the inflammatory response. J Mol Med (Berl) 2009;87:235–47.
- [36] Haitoglou CS, Tsilibary EC, Brownlee M, Charonis AS. Altered cellular interactions between endothelial cells and nonenzymaticallyglucosylatedlaminin/type IV collagen. J Biol Chem 1992;267:12404–7.
- [37] Giacco F, Brownlee M. Oxidative stress and diabetic complications. Circ Res 2010;107:1058–70.
- [38] Ramasamy R, Yan SF, Schmidt AM. Advanced glycation endproducts: from precursors to RAGE: round and round we go. Amino Acids 2012;42:1151–61.
- [39] Yan SF, D'Agati V, Schmidt AM, Ramasamy R. Receptor for Advanced Glycation Endproducts (RAGE): a formidable force in the pathogenesis of the cardiovascular complications of diabetes & aging. Curr Mol Med 2007;7:699–710.
- [40] Raucci A, Cugusi S, Antonelli A, Barabino SM, Monti L, Bierhaus A, et al. A soluble form of the receptor for advanced glycation endproducts (RAGE) is produced by proteolytic cleavage of the membrane-bound form by the sheddase a disintegrin and metalloprotease 10 (ADAM10). FASEB J 2008;22:3716–27.
- [41] Falcone C, Bozzini S, Guasti L, D'Angelo A, Capettini AC, Paganini EM, et al. Soluble RAGE plasma levels in patients with coronary artery disease and peripheral artery disease. ScientificWorldJournal 2013;2013:584504–10.
- [42] Giallauria F, Orio F, Palomba S, Lombardi G, Colao A, Vigorito C. Cardiovascular risk in women with polycystic ovary syndrome. J Cardiovasc Med (Hagerstown) 2008;9:987–92.
- [43] Hoffman LK, Ehrmann DA. Cardiometabolic features of polycystic ovary syndrome. Nat Clin Pract Endocrinol Metab 2008;4:215–22.

- [44] van der Spuy ZM, Dyer SJ. The pathogenesis of infertility and early pregnancy loss in polycystic ovary syndrome. Best Pract Res Clin Obstet Gynaecol 2004;18:755–71.
- [45] Sekar N, Garmey JC, Veldhuis JD. Mechanisms underlying the steroidogenic synergy of insulin and luteinizing hormone in porcine granulosa cells: joint amplification of pivotal sterol-regulatory genes encoding the low-density lipoprotein (LDL) receptor, steroidogenic acute regulatory (stAR) protein and cytochrome P450 side-chain cleavage (P450scc) enzyme. Mol Cell Endocrinol 2000;159:25–35.
- [46] Jonard S, Dewailly D. The follicular excess in polycystic ovaries, due to intra-ovarian hyperandrogenism, may be the main culprit for the follicular arrest. Hum Reprod Update 2004;10:107–17.
- [47] Morley P, Calaresu FR, Barbe GJ, Armstrong DT. Insulin enhances luteinizing hormone-stimulated steroidogenesis by porcine theca cells. Biol Reprod 1989;40:735–43.
- [48] Gonzalez F. Inflammation in Polycystic Ovary Syndrome: underpinning of insulin resistance and ovarian dysfunction. Steroids 2012;77:300–5.
- [49] Taponen S, Martikainen H, Jarvelin MR, Laitinen J, Pouta A, Hartikainen AL, et al. Hormonal profile of women with selfreported symptoms of oligomenorrhea and/or hirsutism: Northern Finland birth cohort 1966 study. J Clin Endocrinol Metab 2003;88:141–7.
- [50] Diamanti-Kandarakis E, Argyrakopoulou G, Economou F, Kandaraki E, Koutsilieris M. Defects in insulin signaling pathways in ovarian steroidogenesis and other tissues in polycystic ovary syndrome (PCOS). J Steroid Biochem Mol Biol 2008;109:242–6.
- [51] Codner E, Escobar-Morreale HF. Clinical review: Hyperandrogenism and polycystic ovary syndrome in women with type 1 diabetes mellitus. J Clin Endocrinol Metab 2007;92:1209–16.
- [52] Nestler JE, Powers LP, Matt DW, Steingold KA, Plymate SR, Rittmaster RS, et al. A direct effect of hyperinsulinemia on serum sex hormone-binding globulin levels in obese women with the polycystic ovary syndrome. J Clin Endocrinol Metab 1991;72:83–9.
- [53] Legro RS, Gnatuk CL, Kunselman AR, Dunaif A. Changes in glucose tolerance over time in women with polycystic ovary syndrome: a controlled study. J Clin Endocrinol Metab 2005; 90:3236–42.
- [54] Diamanti-Kandarakis E, Piperi C, Kalofoutis A, Creatsas G. Increased levels of serum advanced glycation end-products in women with polycystic ovary syndrome. Clin Endocrinol (Oxf) 2005;62:37–43.
- [55] Tantalaki E, Piperi C, Livadas S, Kollias A, Adamopoulos C, Koulouri A, et al. Impact of dietary modification of advanced glycation end products (AGEs) on the hormonal and metabolic profile of women with polycystic ovary syndrome (PCOS). Hormones (Athens) 2014;13:65–73.
- [56] Altieri P, Cavazza C, Pasqui F, Morselli AM, Gambineri A, Pasquali R. Dietary habits and their relationship with hormones and metabolism in overweight and obese women with polycystic ovary syndrome. Clin Endocrinol (Oxf) 2013; 78:52–9.
- [57] Cassese A, Esposito I, Fiory F, Barbagallo AP, Paturzo F, Mirra P, et al. In skeletal muscle advanced glycation end products (AGEs) inhibit insulin action and induce the formation of multimolecular complexes including the receptor for AGEs. J Biol Chem 2008;283:36088–99.
- [58] Ott C, Jacobs K, Haucke E, Navarrete Santos A, Grune T, Simm A. Role of advanced glycation end products in cellular signaling. Redox Biol 2014;2:411–29.
- [59] Pasquali R, Stener-Victorin E, Yildiz BO, Duleba AJ, Hoeger K, Mason H, et al. PCOS Forum: research in polycystic ovary syndrome today and tomorrow. Clin Endocrinol (Oxf) 2011; 74:424–33.

- [60] Dunaif A, Wu X, Lee A, Diamanti-Kandarakis E. Defects in insulin receptor signaling in vivo in the polycystic ovary syndrome (PCOS). Am J Physiol Endocrinol Metab 2001;281: E392-9
- [61] Sano H, Higashi T, Matsumoto K, Melkko J, Jinnouchi Y, Ikeda K, et al. Insulin enhances macrophage scavenger receptor-mediated endocytic uptake of advanced glycation end products. J Biol Chem 1998;273:8630–7.
- [62] Tatone C, Amicarelli F. The aging ovary–the poor granulosa cells. Fertil Steril 2013;99:12–7.
- [63] Diamanti-Kandarakis E, Chatzigeorgiou A, Papageorgiou E, Koundouras D, Koutsilieris M. Advanced glycation endproducts and insulin signaling in granulosa cells. Exp Biol Med (Maywood) 2015 [pii: 1535370215584937, Epub ahead of print].
- [64] Merhi Z, Irani M, Doswell AD, Ambroggio J. Follicular fluid soluble receptor for advanced glycation end-products (sRAGE): a potential indicator of ovarian reserve. J Clin Endocrinol Metab 2014;99:E226–33.
- [65] Merhi Z. Advanced glycation end products and their relevance in female reproduction. Hum Reprod 2014;29:135–45.
- [66] Diamanti-Kandarakis E, Dunaif A. Insulin resistance and the polycystic ovary syndrome revisited: an update on mechanisms and implications. Endocr Rev 2012;33:981–1030.
- [67] Beydoun HA, Beydoun MA, Wiggins N, Stadtmauer L. Relationship of obesity-related disturbances with LH/FSH ratio among post-menopausal women in the United States. Maturitas 2012;71:55–61.
- [68] Magnotti M, Futterweit W. Obesity and the polycystic ovary syndrome. Med Clin North Am 2007;91:1151–68 [ix-x].
- [69] Garruti G, Depalo R, Vita MG, Lorusso F, Giampetruzzi F, Damato AB, et al. Adipose tissue, metabolic syndrome and polycystic ovary syndrome: from pathophysiology to treatment. Reprod Biomed Online 2009;19:552–63.
- [70] Teede HJ, Joham AE, Paul E, Moran LJ, Loxton D, Jolley D, et al. Longitudinal weight gain in women identified with polycystic ovary syndrome: results of an observational study in young women. Obesity (Silver Spring) 2013;21:1526–32.
- [71] Stepto NK, Cassar S, Joham AE, Hutchison SK, Harrison CL, Goldstein RF, et al. Women with polycystic ovary syndrome have intrinsic insulin resistance on euglycaemichyperinsulaemic clamp. Hum Reprod 2013;28:777–84.
- [72] Diamanti-Kandarakis E, Piouka A, Livadas S, Piperi C, Katsikis I, Papavassiliou AG, et al. Anti-mullerian hormone is associated with advanced glycosylated end products in lean women with polycystic ovary syndrome. Eur J Endocrinol 2009;160:847–53.
- [73] Cai W, Ramdas M, Zhu L, Chen X, Striker GE, Vlassara H. Oral advanced glycation endproducts (AGEs) promote insulin resistance and diabetes by depleting the antioxidant defenses AGE receptor-1 and sirtuin 1. Proc Natl Acad Sci U S A 2012;109:15888–93.
- [74] Uribarri J, Cai W, Ramdas M, Goodman S, Pyzik R, Chen X, et al. Restriction of advanced glycation end products improves insulin resistance in human type 2 diabetes: potential role of AGER1 and SIRT1. Diabetes Care 2011;34:1610–6.
- [75] Mark AB, Poulsen MW, Andersen S, Andersen JM, Bak MJ, Ritz C, et al. Consumption of a diet low in advanced glycation end products for 4 weeks improves insulin sensitivity in overweight women. Diabetes Care 2014;37:88–95.
- [76] Vlassara H, Striker GE. AGE restriction in diabetes mellitus: a paradigm shift. Nat Rev Endocrinol 2011;7:526–39.
- [77] Diamanti-Kandarakis E, Alexandraki K, Piperi C, Aessopos A, Paterakis T, Katsikis I, et al. Effect of metformin administration on plasma advanced glycation end product levels in women with polycystic ovary syndrome. Metabolism 2007;56:129–34.
- [78] Li M, Yang M, Zhou X, Fang X, Hu W, Zhu W, et al. Elevated circulating levels of irisin and the effect of metformin treatment in women with polycystic ovary syndrome. J Clin Endocrinol Metab 2015;100:1485–93.

- [79] Masha A, Manieri C, Dinatale S, Bruno GA, Ghigo E, Martina V. Prolonged treatment with N-acetylcysteine and L-arginine restores gonadal function in patients with polycystic ovary syndrome. J Endocrinol Invest 2009;32:870–2.
- [80] Kahal H, Aye M, Rigby AS, Sathyapalan T, England RJ, Atkin SL. The effect of parathyroidectomy on neuropsychological symptoms and biochemical parameters in patients with asymptomatic primary hyperparathyroidism. Clin Endocrinol (Oxf) 2012;76:196–200.
- [81] Jia X, Chang T, Wilson TW, Wu L. Methylglyoxal mediates adipocyte proliferation by increasing phosphorylation of Akt1. PLoS One 2012;7:e36610–8.
- [82] Uribarri J, Cai W, Woodward M, Tripp E, Goldberg L, Pyzik R, et al. Elevated serum advanced glycation endproducts in obese indicate risk for the metabolic syndrome: a link between healthy and unhealthy obesity? J Clin Endocrinol Metab 2015;100:1957–66.
- [83] Teillet L, Verbeke P, Gouraud S, Bakala H, Borot-Laloi C, Heudes D, et al. Food restriction prevents advanced glycation end product accumulation and retards kidney aging in lean rats. J Am Soc Nephrol 2000;11:1488–97.
- [84] Gugliucci A, Kotani K, Taing J, Matsuoka Y, Sano Y, Yoshimura M, et al. Short-term low calorie diet intervention reduces serum advanced glycation end products in healthy overweight or obese adults. Ann Nutr Metab 2009;54:197–201.
- [85] Gaens KH, Goossens GH, Niessen PM, van Greevenbroek MM, van der Kallen CJ, Niessen HW, et al. Nepsilon-(carboxymethyl)lysine-receptor for advanced glycation end product axis is a key modulator of obesity-induced dysregulation of adipokine expression and insulin resistance. Arterioscler Thromb Vasc Biol 2014;34:1199–208.
- [86] Yamamoto Y, Yamamoto H. RAGE-Mediated Inflammation, Type 2 Diabetes, and Diabetic Vascular Complication. Front Endocrinol (Lausanne) 2013;4:105–7.
- [87] Ueno H, Koyama H, Shoji T, Monden M, Fukumoto S, Tanaka S, et al. Receptor for advanced glycation end-products (RAGE) regulation of adiposity and adiponectin is associated with atherogenesis in apoE-deficient mouse. Atherosclerosis 2010; 211:431–6.
- [88] He CT, Lee CH, Hsieh CH, Hsiao FC. Soluble form of receptor for advanced glycation end products is associated with obesity and metabolic syndrome in adolescents. Int J Endocrinol 2014;2014:657607–13.
- [89] Davis KE, Prasad C, Vijayagopal P, Juma S, Imrhan V. Serum soluble receptor for advanced glycation end products correlates inversely with measures of adiposity in young adults. Nutr Res 2014;34:478–85.
- [90] Brix JM, Hollerl F, Kopp HP, Schernthaner GH, Schernthaner G. The soluble form of the receptor of advanced glycation endproducts increases after bariatric surgery in morbid obesity. Int J Obes (Lond) 2012;36:1412–7.
- [91] Phelan N, O'Connor A, Kyaw Tun T, Correia N, Boran G, Roche HM, et al. Hormonal and metabolic effects of polyunsaturated fatty acids in young women with polycystic ovary syndrome: results from a cross-sectional analysis and a randomized, placebo-controlled, crossover trial. Am J Clin Nutr 2011;93:652–62.
- [92] Rafraf M, Mohammadi E, Asghari-Jafarabadi M, Farzadi L. Omega-3 fatty acids improve glucose metabolism without effects on obesity values and serum visfatin levels in women with polycystic ovary syndrome. J Am Coll Nutr 2012;31:361–8.
- [93] Mak W, Dokras A. Polycystic ovarian syndrome and the risk of cardiovascular disease and thrombosis. Semin Thromb Hemost 2009;35:613–20.
- [94] Karoli R, Fatima J, Siddiqi Z, Vatsal P, Sultania AR, Maini S. Study of early atherosclerotic markers in women with polycystic ovary syndrome. Indian J Endocrinol Metab 2012; 16:1004–8.

- [95] Orio Jr F, Palomba S, Cascella T, Manguso F, Vuolo L, Tafuri D, et al. Early impairment of endothelial structure and function in young normal-weight women with polycystic ovary syndrome. J Clin Endocrinol Metab 2004;89:4588–93.
- [96] Guleria AK, Syal SK, Kapoor A, Kumar S, Tiwari P, Dabadghao P. Cardiovascular disease risk in young Indian women with polycystic ovary syndrome. Gynecol Endocrinol 2014;30:26–9.
- [97] Teng HW, Chien YW, Hsu MI, Chen CI. The relationship between carotid intima-media thickness and endogenous androgens in young women with polycystic ovary syndrome in Taiwan. Gynecol Endocrinol 2013;29:238–41.
- [98] Sprung VS, Atkinson G, Cuthbertson DJ, Pugh CJ, Aziz N, Green DJ, et al. Endothelial function measured using flow-mediated dilation in polycystic ovary syndrome: a meta-analysis of the observational studies. Clin Endocrinol (Oxf) 2013;78:438–46.
- [99] Rajendran S, Willoughby SR, Chan WP, Liberts EA, Heresztyn T, Saha M, et al. Polycystic ovary syndrome is associated with severe platelet and endothelial dysfunction in both obese and lean subjects. Atherosclerosis 2009;204:509–14.
- [100] Lambert EA, Teede H, Sari CI, Jona E, Shorakae S, Woodington K, et al. Sympathetic activation and endothelial dysfunction in polycystic ovary syndrome are not explained by either obesity or insulin resistance. Clin Endocrinol (Oxf) 2015. http://dx.doi.org/10.1111/cen.12803 [Epub ahead of print].
- [101] Rizvi AA. Cytokine biomarkers, endothelial inflammation, and atherosclerosis in the metabolic syndrome: emerging concepts. Am J Med Sci 2009;338:310–8.
- [102] Chan WP, Ngo DT, Sverdlov AL, Rajendran S, Stafford I, Heresztyn T, et al. Premature aging of cardiovascular/ platelet function in polycystic ovarian syndrome. Am J Med 2013;126:640e1–7.
- [103] Corti R, Hutter R, Badimon JJ, Fuster V. Evolving concepts in the triad of atherosclerosis, inflammation and thrombosis. J Thromb Thrombolysis 2004;17:35–44.
- [104] Diamanti-Kandarakis E, Alexandraki K, Piperi C, Protogerou A, Katsikis I, Paterakis T, et al. Inflammatory and endothelial markers in women with polycystic ovary syndrome. Eur J Clin Invest 2006:36:691–7.
- [105] Christakou C, Economou F, Livadas S, Piperi C, Adamopoulos C, Marinakis E, et al. Strong and positive association of endothelin-1 with AGEs in PCOS: a causal relationship or a bystander? Hormones (Athens) 2011;10:292–7.
- [106] Kocer D, Bayram F, Diri H. The effects of metformin on endothelial dysfunction, lipid metabolism and oxidative stress in women with polycystic ovary syndrome. Gynecol Endocrinol 2014;30:367–71.
- [107] Piarulli F, Lapolla A, Ragazzi E, Susana A, Sechi A, Nollino L, et al. Role of endogenous secretory RAGE (esRAGE) in defending against plaque formation induced by oxidative stress in type 2 diabetic patients. Atherosclerosis 2013;226:252–7.
- [108] Lu L, Pu LJ, Zhang Q, Wang LJ, Kang S, Zhang RY, et al. Increased glycated albumin and decreased esRAGE levels are related to angiographic severity and extent of coronary artery disease in patients with type 2 diabetes. Atherosclerosis 2009;206:540–5.
- [109] Peng WH, Lu L, Hu J, Yan XX, Zhang Q, Zhang RY, et al. Decreased serum esRAGE level is associated with angiographically determined coronary plaque progression in diabetic patients. Clin Biochem 2009;42:1252–9.

- [110] Tanaka N, Yonekura H, Yamagishi S, Fujimori H, Yamamoto Y, Yamamoto H. The receptor for advanced glycation end products is induced by the glycation products themselves and tumor necrosis factor-alpha through nuclear factor-kappa B, and by 17beta-estradiol through Sp-1 in human vascular endothelial cells. J Biol Chem 2000;275:25781–90.
- [111] Escobar-Morreale HF, Calvo RM, Sancho J, San Millan JL. TNF-alpha and hyperandrogenism: a clinical, biochemical, and molecular genetic study. J Clin Endocrinol Metab 2001; 86:3761–7.
- [112] Yun JH, Choi JW, Lee KJ, Shin JS, Baek KH. The promoter -1031(T/C) polymorphism in tumor necrosis factor-alpha associated with polycystic ovary syndrome. Reprod Biol Endocrinol 2011;9:131–6.
- [113] Pepene CE, Ilie IR, Marian I, Duncea I. Circulating osteoprotegerin and soluble receptor activator of nuclear factor kappaB ligand in polycystic ovary syndrome: relationships to insulin resistance and endothelial dysfunction. Eur J Endocrinol 2011;164:61–8.
- [114] Toulis KA, Goulis DG, Mintziori G, Venetis CA, Kolibianakis EM, Tarlatzis BC. Meta-analysis of cardiovascular disease risk markers in women with polycystic ovary syndrome. Hum Reprod Update 2011;17:741–60.
- [115] Siroen MP, Teerlink T, Nijveldt RJ, Prins HA, Richir MC, van Leeuwen PA. The clinical significance of asymmetric dimethylarginine. Annu Rev Nutr 2006;26:203–28.
- [116] Raptis V, Kapoulas S, Grekas D. Role of asymmetrical dimethylarginine in the progression of renal disease. Nephrology (Carlton) 2013;18:11–21.
- [117] Das UN, Repossi G, Dain A, Eynard AR. L-arginine, NO and asymmetrical dimethylarginine in hypertension and type 2 diabetes. Front Biosci (Landmark Ed) 2011;16: 13–20.
- [118] Boger RH, Bode-Boger SM, Szuba A, Tsao PS, Chan JR, Tangphao O, et al. Asymmetric dimethylarginine (ADMA): a novel risk factor for endothelial dysfunction: its role in hypercholesterolemia. Circulation 1998;98:1842–7.
- [119] Lundman P, Eriksson MJ, Stuhlinger M, Cooke JP, Hamsten A, Tornvall P. Mild-to-moderate hypertriglyceridemia in young men is associated with endothelial dysfunction and increased plasma concentrations of asymmetric dimethylarginine. J Am Coll Cardiol 2001;38:111–6.
- [120] Bode-Boger SM, Scalera F, Ignarro LJ. The L-arginine paradox: Importance of the L-arginine/asymmetrical dimethylarginine ratio. Pharmacol Ther 2007;114:295–306.
- [121] Ngo DT, Chan WP, Rajendran S, Heresztyn T, Amarasekera A, Sverdlov AL, et al. Determinants of insulin responsiveness in young women: Impact of polycystic ovarian syndrome, nitric oxide, and vitamin D. Nitric Oxide 2011;25:326–30.
- [122] Turkcuoglu I, Engin-Ustun Y, Turan F, Kali Z, Karabulut AB, Meydanli M, et al. Evaluation of asymmetric dimethylarginine, nitric oxide levels and associated independent variables in obese and lean patients with polycystic ovarian syndrome. Gynecol Endocrinol 2011;27:609–14.
- [123] Heutling D, Schulz H, Nickel I, Kleinstein J, Kaltwasser P, Westphal S, et al. Asymmetrical dimethylarginine, inflammatory and metabolic parameters in women with polycystic ovary syndrome before and after metformin treatment. J Clin Endocrinol Metab 2008;93:82–90.